



Genetic Aspects of Acromegaly

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Abstract

The prevalence of acromegaly is 40 to 70 cases per million with an annual incidence of 3 to 4 new cases per million of population. Patients with pituitary tumors usually do not have a family history of the pituitary gland diseases, and in most cases, embryo or mosaic changes cannot be detected, while recurrent somatic mutations are common in ubiquitin specific peptidase 8, in corticotropinomas and guanine nucleotide binding protein, alpha stimulating complex locus in somatotropinomas genes. The hereditary pituitary tumors are rare (~5%), but are currently well known and represented as a family disease or manifested sporadic cases. Sporadic cases can have mutations of the germline *de novo* (for example, most cases of X-linked acrogigantism due to duplication of GPR, mosaic mutations (McCune-Albright syndrome [GNAS mutations] or XLAG) whereas family history is masked as low penetrance (aryl hydrocarbon receptor-interacting protein, AIP) or imprinting.

Introduction

The pituitary tumors are the most common intracranial neoplasms, affecting 1/1000 population of the world [1]. Although, they are usually benign, the adenomas of the anterior pituitary gland cause excess or deficiency of hormones, tumor growth and the need for neurosurgical, radiological and/or long-term medical therapy to control the disease. Adenomas that secrete Growth Hormone (GH), also known as somatotropinomas, make up about 20% of all the pituitary tumors and are the second most common pituitary tumor, securing the hormone, after tumors securing Prolactin (PRL), which accounts for 40% to 45% of all pituitary tumors [2,3]. The prevalence of acromegaly is 40 to 70 cases per million with an annual incidence of 3 to 4 new cases per million of population [4-6]. Patients with pituitary tumors usually do not have a family history of the pituitary gland diseases, and in most cases, embryo or mosaic changes cannot be detected, while recurrent somatic mutations are common in Ubiquitin Specific Peptidase 8 (USP8), in corticotropinomas [7-14] and guanine nucleotide binding protein, alpha stimulating complex locus (GNAS) in somatotropinomas genes. The hereditary pituitary tumors are rare (~5%), but are currently well known and represented as a family disease or manifested sporadic cases. Sporadic cases can have mutations of the germline *de novo* (for example, most cases of X-Linked Acrogigantism (XLAG) due to duplication of GPR101, mosaic mutations (McCune-Albright syndrome [GNAS mutations] or XLAG) whereas family history is masked as low penetrance (Aryl hydrocarbon receptor-Interacting Protein, AIP) or imprinting (SDHD) (Table 1). Genetic defects can predispose to the development of a pituitary tumor either as an isolated disease (AIP, GPR101), or as part of the syndrome predisposing to the tumor: Multiple Endocrine Neoplasia type 1 (MEN1), type 4 (CDKN 1B) or other Cyclin-Dependent Kinase Inhibitor (CDKN 1A, CDKN 2B or CDKN 2C), type 5 (MAX) [15,16], pituitary tumors, pheochromocytomas or paraganglioma (SDHx), Carney complex (PRKAR1A) and the potential risk of the pituitary adenoma development with Lynch syndrome (MSH2 and PMS2) [17,18]. Genes with germline mutations that predispose the syndrome to tumors can also contribute to the tumor pathogenesis of sporadic tumors through somatic changes. However, this rarely happens to the genes associated with the pituitary tumor. GNAS is often found in the form of a somatic mutation in tumor tissue and in the form of a mosaic mutation in McCune-Albright syndrome. GPR101 is considered as an embryo or mosaic mutation. Understanding the genetic background of the pituitary tumors can lead to an early diagnosis of the disease. Moreover, an understanding of their molecular mechanism can also open opportunities for targeted medical therapy even for sporadic tumors.

Embryo or Mosaic Mutations

Patients with mutations of the germline associated with the pituitary tumors are classified as isolated pituitary tumors. The first group includes relatives with AIP and GPR101 germs and,

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possibly, recently described by changes in Cdk5 and Abl Enzyme Substrate 1 (CABLES1), in addition to a large number of Familial Isolated Pituitary Adenomas (FIPA) without identified genetic anomalies, while the last group includes MEN1, MEN4 and recently discovered MEN5, the Carney complex, paragangliomas, diseases associated with related genes (SDHx), DICER1, Lynch and McCune-Albright syndromes. The general term "Familial Isolated Pituitary Adenomas (FIPA)" refers to a family consisting of at least two family members with pituitary tumors in the absence of other associated tumors [19,20]. In most families with FIPA (90%) there is no genetic mutation can be identified among major genes predisposing to pituitary adenoma. On the other hand, mutations of the embryo line can also be identified in sporadic cases (so-called simple cases) due to *de novo* mutations, as is often the case with XLAG syndrome, or due to low penetrance and, therefore, an inconspicuous family history, as is usually observed in cases with AIP mutation, or simply the lack of information among family members about the disease. FIPA is a heterogeneous state with significant differences in the phenotype. While homogeneous (the same type of tumor for all family members) and heterogeneous (different types of tumors for family members) cases can be distinguished, this differentiation complicated with the common somatomammotroph and other multihormonal or clinically asymptomatic hormones producing phenotypes.

Aryl Hydrocarbon Receptor-Interacting Protein

Embryonic heterozygous inactivating mutations in AIP gene are present in about 10% of FIPA families [19] and demonstrate an autosomal-dominant type of inheritance with incomplete penetrance. About half of the families with isolated GH-producing tumors have mutations in this gene [21]. The AIP gene encodes a koseperon of 330 amino acids involved in many processes, including subcellular transfer, stability of nuclear receptors and transactivation potential [19-23]. It is assumed that in the AIP pituitary gland acts as a tumor suppressor, which is confirmed by the fact that mutations are represented with the function failure and the presence of a normal copy of the gene (Loss of Heterozygosity, LOH, usually due to the loss of part of the chromosome area 11q [13] in pituitary tumors with AIP mutation. In patients with sporadic pituitary macroadenomas younger than 18 years-old AIP germline mutations can be found in 20% of cases (only 3% in cases of unremarkable sporadic pituitary tumors). Patients with AIP mutation usually have somatotropinomas mixed with somatomammotropinoma or prolactinomas (10%), rarely with clinically asymptomatic somatotropinomas and a single case of thyrotropinoma. There are also several cases of somatotrophic hyperplasia with or without associated tumors. During prospective screening, patients with clinically non-functioning microadenomas were identified, in which pathology is unknown. The presence of AIP mutations in patients is usually associated with macroadenomas in young age, usually with invasive pattern and a predisposition to apoplexy and poor response to the first-generation somatostatin analogues, while second-generation somatostatin analogues showed a good response in some cases [21,24]. Thus, there is a spectrum of manifestations of patients with positive AIP mutations, ranging from disease manifestation period up to five years to cases when the disease is not diagnosed up to an older age, while an aggressive disease in adolescence is the most typical clinical manifestation of penetrant cases. AIP, as a chaperon, has several interacting partners. While the aryl hydrocarbon receptor (AIP) gave the name to the protein, and their interaction was studied in relation to the pituitary

oncogenesis, the causal association has not yet been shown. AIP is a transcription factor activated by a ligand, whose client proteins include Phosphodiesterase type 4 (PDE4), which switches off the cAMP path. AIP is expressed in somatotropinomas and NFPA, and AIP polymorphism was associated with aggressive tumor behavior with transformation into sporadic somatotropinomas [25,26]. Molecular factors causing a decrease in the response to the first-generation somatostatin analogues were widely discussed [27-29]: Low level of type 2 Somatostatin Receptor (SSRT2), [30] high level of expression of 5TMD4 depression [31] and low ratio of SSRT2/SSRT5 [32] or the absence of ZAC1 [33,34]. On plasma membrane, AIP plays an important role in the initial stages of the RET-apoptosis path in cells expressing PIT1. The absence of AIP or pathogenic mutations blocks this path, contributing to the RET-survival characteristic of the pituitary tumors [35]. This mechanism can also explain the unique tissue and cellular specificity of tumors associated with AIP, since PIT1 and RET are usually expressed only in somato- and lactotrophic cells. It was found that in tumors with a positive AIP, microRNA-34a (miR-34a) is activated, which correlates with prooncogenic characteristics, high levels of cAMP and a wrong response to first-generation somatostatin analogues [36-39].

Family FIPA without Common Mutations

Most (90%) family FIPA do not have known genetic mutations [21]. Although the type of inheritance can be compatible with monogenic autosomal dominant variant with very low penetrance, an oligogenic inheritance model is more likely. In the cohort of FIPA International Consortium of 318 AIP-negative (and GPR101) FIPA 21% have homogeneous acromegaly (which is 46% of 146 homogeneous AIP-negative families), and 32% have heterogeneous FIPA, at least in one family with acromegaly (54% of 172 heterogeneous families with the absence of AIP mutations) [21]. Although, somatotropinoma is not as common as in patients with a positive AIP mutation, somatotropinoma is the most common subtype in patients with a negative AIP mutation. Among heterogeneous groups (172 kinds), 29% have acromegaly and prolactinoma, 26% have acromegaly and NFPA, 4% have acromegaly and Cushing's disease [21]. In these families, only two or three patients with this disease are usually identified. The disease very rarely begins in childhood (≤ 18 years), but more often in adulthood (the average age at the time of diagnosis in the cohort of patients with AIP-negative FIPA is 38 years) [40]. It is impossible to establish accurate penetrance among AIP-negative FIPA. When comparing the AIP-positive and AIP-negative FIPA, the number of affected cases is much lower in AIP-negative persons, which implies lower penetrance [41].

XLAG (GPR101)

X-Linked Acrogigantism Syndrome (XLAG) is a rare condition performed with acrogigantism, manifested in early childhood. Hypophyseal gigantism and clinical signs of acromegaly occur in children during the first few years of life due to excessively high levels of GH and IGF-1 [40-42]. This disease has 100% penetrance and is considered one of two confirmed genetic causes of FIPA [42]. XLAG is caused by the germinal or somatic mosaic duplication of the GPR101 gene, leading to the effect of accelerating the function [43-46] GPR101 (located on the XQ26.3) codes the receptor associated with the G-protein, and is a strong activator for cAMP path [42-49]. GPR101 duplications are more common in women (80% of cases, this is more expected 66% due to the gene located in the X-chromosome) as a mutation of the germline *de novo*. Most men with highest height

Table 1: Genetic conditions with GH secretin pituitary adenomas.

Gene name	Mutation of the embryo line	Mosaic mutation	Somatic mutations in pituitary	Embryo line: isolated or syndromal	Notice
AIP	Yes	?	No	isolated	Penetrance 20%–23%
ATRXa	No	?	Yes	somatic	The biallel embryo line causes a neurological disorder. Somatic changes in aggressive pituitary tumors. Somatic mutations are also present in other neuroendocrine tumors.
CABLES1a	Yes	?	?	Isolated	Corticotrophic tumors, all mutations were Missense and not one of them was shortened. There are no family cases
CDH23a	Yes	?	?	Isolated	CDH23 homozygous mutations are associated with the deafness and Asher syndrome. There are no reports of patients with the pituitary adenoma in these families.
CDKN1Aa	Yes	?	?	syndromal	MEN1/4-like syndrome
CDKN1Ba	Yes	?	Yes (rare)	syndromal	Some cases of corticotrophic tumor in children without well -known other syndromic manifestations
CDKN2Ba	Yes	?	?	syndromal	MEN1- like syndrome
CDKN2Ca	Yes	?	Yes (rare)	syndromal	MEN1- like syndrome
DICER1	Yes	Yes	Yes	Syndromal	The blastoma of the pituitary gland with the early beginning, usually secreting ACTH, one teenage case
GNAS	No	Yes	Yes	Syndromal	Mosaic (McCune-Albright syndrome) or somatic (30–40% of somatotroph adenomas)
GPR101	Yes	Yes	No	Isolated	Data refers to genes duplication
MAXa	Yes	?	?	Syndromal	Several cases with MAX large deletions
MEN1	Yes	Yes	Yes	syndromal	Penetrance 40%
MLH1a	Yes	?	?	syndromal	Lynch syndrome, one case of pituitary carcinoma
MSH2a	Yes	?	?	syndromal	Lynch syndrome, aggressive corticotroph and lactotroph pituitary tumors
MSH6a	Yes	?	?	syndromal	Lynch syndrome, mainly corticotroph pituitary tumors
NF1b	Yes	?	?	syndromal	Excess growth hormone in 10% of patients with optic nerve glioma not related to pituitary tumors
NR3C1a	No	?	Yes	syndromal	The mutation of the embryo line usually causes resistance syndrome to glucocorticoids
PMS2a	Yes	?	?	syndromal	Lynch syndrome, mainly corticotroph pituitary tumors.
PRKAR1A	Yes	Yes	No	syndromal	Large deletions of PRKAR1A lead to a more severe phenotype of the disease
PRKACAa	Yes	?	?	syndromal	Single case
SDHx	Yes	?	Yes (rare)	syndromal	SDHX mutations can be associated with the tumors of the pituitary gland, but the penetrance is very low

[50] have somatic mosaicism, while in three described families the transfer was from mother to son. The pituitary tumors (95%) with significant co-secretion of prolactin, are not as aggressive as in patients with AIP mutations, and in 25% of cases the cause of the disease is the pituitary hyperplasia, not a tumor [42,46-50].

Multiple Endocrine Neoplasia Type 1 (MEN1)

MEN1 is a tumor disease affecting multiple organs with three main manifestations: Neoplasms of parathyroid glands that cause primary hyperparathyroidism, neuroendocrine pancreatic tumors and approximately 30% to 40% of the pituitary tumors, also, thymus, skin and adrenal tumors [51-54]. MEN1 is diagnosed clinically if (1) the patient has two or more tumors associated with MEN1; (2) a patient has a relative of the first degree of kinship with MEN1 and one tumor associated with MEN1, or; (3) the carrier of the mutant gene with the MEN1 mutation, but without clinical, biochemical or anatomical features of MEN1 [55]. The main genetic defect is inherited by an autosomal-dominant type of mutation with a loss of function in MEN1 gene, usually accompanied by a second change in another allele at a somatic level [56]. Most cases develop either in families (up to 90%) or as *de novo* mutations (10% of cases) [57,58] and although in general, the syndrome demonstrates complete penetrance with biochemical changes present by the fifth decade of lifetime in more than 95% of patients [59], the pituitary aspect has ~40% penetrance.

Gene MEN1 (11Q13) encodes menin, a tumor-protein, which plays an important role in the regulation of transcription, genome stability, cellular division and control of the cell cycle. The pituitary tumors are the first feature of approximately 15% to 20% of cases of family MEN1 patients who are usually found in women with an average age of 40 years. There is a small inconsistency in the types of pituitary tumors associated with MEN1, depending on how the data was collected. In a cohort with clinical manifestations, prolactinomas are the most common lesions (60%), usually developing in childhood, whereas acromegaly contributes 25%, NFPA (15%), corticotropes (5%). In prospective cohort prolactinomas make up to 42%, 42% of NFAPs (many of them have only a few millimeters size and are difficult to differentiate from incidentaloma), 7% of acromegaly and mixed tumors and corticotropes [60-65]. The sporadic tumors of the pituitary gland with the somatic mutation of MEN1 are rare (3.5%), but menin levels were decreased in most pituitary tumors and were absent in metastatic pituitary carcinoma [60].

MEN4

MEN4 [66,67] have a MEN1-like phenotype with primary hyperparathyroidism and pituitary tumors (prolactinomas, somatotropinomas, corticotropinomas and NFPA) found in approximately 40% of patients, but with functionally intact genome of MEN1 [68,69]. In carriers of MEN4 mutations found during family

screening, the NFPA was described as the most common pituitary tumor [69-71]. Due to the rarity of the disease, the penetrance and correlation of the genotype-phenotype evaluation chance is poor. These pituitary tumors have an average age when diagnosed from 30 to 79 years, are usually not aggressive and are treated well [69]. While MEN4 was first observed in rats with MEN1- and MEN2-like tumors [72,73] due to the mutation in the inhibitor of the P27 cell cycle (codes CDKN1b), other mutations of the cell cycle inhibitor (CDKN2B (P15), CDKN2C (P18) and CDKN1A (P21) [74]. CDKN1B also found in several cases with corticotropinomas in children [74,75].

Carney Complex

The Carney complex is a rare heterozygous autosomal dominant disease with multiple endocrine and non-endocrine symptoms, including pituitary hyperplasia or tumor with complete penetrance [76,77]. The disorder can be diagnosed clinically if there are two or more basic criteria or one large criterion and there is a patient relative of the first degree of kinship or a well-known inactivating mutation of PRKAR1A. The main criteria are spotted skin hyperpigmentation, skin and cardiac myxomas, Primary Pigmented Nodular Adrenocortical Disease (PPNAD), acromegaly, large cell calcifying Sertoli cell tumor of testis, thyroid carcinoma or multiple nodules, mammary gland adenoma, psammomatous melanotic schwannoma, blue naevus and osteochondromyxoma [78,79]. Large deletions lead to a more serious illness, and *de novo* mutations are present in a significant minority of simple cases [80,81]. The PRKAR1A gene (17Q23-24) encodes the PKA regulatory protein of subunit of type 1 α and functions as a tumor suppressor gene [82]. Although most (~70%) patients have an abnormal dynamic of the GH axis and an increased IGF-1 level, only 10% has active acromegaly, often with hyperprolactinemia, usually manifested for the third decade of life [79,83]. Corticotrophic adenomas are rarely described [84,85] and are a diagnostic problem, since the adrenal gland Cushing's syndrome is a common manifestation [86-91].

McCune-Albright Syndrome

McCune-Albright syndrome is a rare genetic disease manifested by fibrous dysplasia, "café-au-lait" skin pigmentation and premature puberty [92-96]. Other endocrine problems include an excess of GH, hypercortisolemia due to nodular adrenal hyperplasia and thyrotoxicosis, but recently pancreatic and hepatic tumors are also described [97-100]. Hypersecretion of GH is observed in 20% of patients with McCune-Albright syndrome, but only about 50% of patients have a pituitary tumor or pituitary hyperplasia discovered on imaging studies. The disorder is caused by somatic activating mutations in the GNAS gene, which occur at the early postzygotic stage, which leads to the somatic mosaicism of this mutation. GNAS encodes guanine-nucleotide-binding protein, α -stimulating polypeptide, and a mutation leads to an increase in the function with constantly increased levels of adenylate cycles, cAMP and PKA activity, which leads to persistent hypersecretion of GH and cell proliferation. Due to the variable level of mosaicism, the degree of phenotype covers a wide range of severity.

Conclusion

Although there was significant progress in the characterization of the molecular basis of the pituitary oncogenesis, current data indicate that any of these changes are hardly responsible for sporadic tumors that most cases account for. In conclusion, genetic testing in the treatment of pituitary diseases is recommended in rare cases.

Understanding the genetic basis of the disease helps to identify patients and family members at risk, facilitates early diagnosis and, therefore, improves the long-term outcomes, and also leads to a better understanding of its oncogenesis.

References

1. Aflorei ED, Korbonits M. Epidemiology and etiopathogenesis of pituitary adenomas. *J Neurocol.* 2014;117:379-94.
2. Agustsson TT, Baldvinsdottir T, Jonasson JG. The epidemiology of pituitary adenomas in Iceland, 1955-2012: A nationwide population based study. *Eur J Endocrinol.* 2015;173:655-64.
3. Herman V, Fagin J, Gonsky R, Kovacs K, Melmed S. Clonal origin of pituitary adenomas. *J Clin Endocrinol Metab.* 1990;71:1427-33.
4. Ostrom QT, Gittleman H, Truitt G, Boscia A, Kruchko C, Barnholtz-Sloane JS. CBTRUS statistical report: Primary brain and other central nervous system tumors diagnosed in the United States in 2011- 2015. *Neuro Oncol.* 2018;20:iv1-iv86.
5. Daly AF, Rixhon M, Adam C, Dempegioti A, Tichomirowa MA, Beckers A. High prevalence of pituitary adenomas: A cross-sectional study in the province of Liege, Belgium. *J Clin Endocrinol Metab.* 2006;91:4769-75.
6. Fernandez A, Karavitaki N, Wass JA. Prevalence of pituitary adenomas: A community-based, cross-sectional study in Banbury (Oxfordshire, UK). *Clin Endocrinol (Oxf).* 2010;72:377-82.
7. Fontana E, Gaillard R. [Epidemiology of pituitary adenoma: Results of the first Swiss study]. *Rev Med Suisse.* 2009;5:2172-4.
8. Gruppeta M, Mercieca C, Vassallo J. Prevalence and incidence of pituitary adenomas: A population based study in Malta. *Pituitary.* 2013;16:545-53.
9. Raappana A, Koivukangas J, Ebeling T, Pirila T. Incidence of pituitary adenomas in Northern Finland in 1992-2007. *J Clin Endocrinol Metab.* 2010;95:4268-75.
10. Molitch ME. Diagnosis and treatment of pituitary adenomas: A review. *JAMA.* 2017;317:516-24.
11. Raverot G, Ilie MD, Lasolle H. Aggressive pituitary tumors and pituitary carcinomas. *Nat Rev Endocrinol.* 2021;17:671-84.
12. Asa SL, Casar-Borota O, Chanson P. From pituitary adenoma to pituitary neuroendocrine tumor (PitNET): An International Pituitary Pathology Club proposal. *Endocr Relat Cancer.* 2017;24:C5-C8.
13. Reincke M, Sbierra S, Hayakawa A. Mutations in the deubiquitinase gene USP8 cause Cushing's disease. *Nat Genet.* 2015;47:31-38.
14. Ma ZY, Song ZJ, Chen JH. Recurrent gain-of-function USP8 mutations in Cushing's disease. *Cell Res.* 2015;25:306-17.
15. Spada A, Vallar L. G-protein oncogenes in acromegaly. *Horm Res.* 1992;38:90-93.
16. Seabrook AJ, Harris JE, Velosa SB. Multiple endocrine tumors associated with germline MAX mutations: multiple endocrine neoplastic type 5? *J Clin Endocrinol Metab.* 2021;106:1163-82.
17. Loughrey PB, Baker G, Herron B. Invasive ACTH-producing pituitary gland neoplasm secondary to MSH2 mutation. *Cancer Genet.* 2021;256-257:36-39.
18. Bengtsson D, Joost P, Aravidis C. Corticotroph pituitary carcinoma in a patient with Lynch Syndrome (LS) and pituitary tumors in a nationwide LS cohort. *J Clin Endocrinol Metab.* 2017;102:3928-32.
19. Beckers A, Aaltonen LA, Daly AF, Karhu A. Familial Isolated Pituitary Adenomas (FIPA) and the pituitary adenoma predisposition due to mutations in the aryl hydrocarbon receptor interacting protein (AIP) gene. *Endocr Rev.* 2013;34:239-77.
20. Daly AF, Jaffrain-Rea ML, Ciccarella A. Clinical characterization of familial

- isolated pituitary adenomas. *J Clin Endocrinol Metab.* 2006;91:3316-23.
21. Marques P, Caimari F, Hernández-Ramírez LC. Significant benefits of AIP testing and clinical screening in familial isolated and young-onset pituitary tumors. *J Clin Endocrinol Metab.* 2020;105:e2247-e2260.
 22. Trivellin G, Korbonits M. AIP and its interacting partners. *J Endocrinol.* 2011;210:137-55.
 23. Sun D, Stopka-Farooqui U, Barry S. Aryl hydrocarbon receptor interacting protein maintains germinal center B cells through suppression of BCL6 degradation. *Cell Rep.* 2019;27:1461-71.e4.
 24. Daly AF, Rostomyan L, Betae D. AIP-mutated acromegaly resistant to first-generation somatostatin analogs: long-term control with pasireotide LAR in two patients. *Endocr Connect.* 2019;8:367-77.
 25. Jaffrain-Rea ML, Angelini M, Gargano D. Expression of Aryl Hydrocarbon Receptor (AHR) and AHR-interacting protein in pituitary adenomas: Pathological and clinical implications. *Endocr Relat Cancer.* 2009;16:1029-43.
 26. Heliövaara E, Raitila A, Launonen V. The expression of AIP related molecules in elucidation of cellular pathways in pituitary adenomas. *Am J Pathol.* 2009;175:2501-7.
 27. Larkin S, Reddy R, Karavitaki N, Cudlip S, Wass J, Ansorge O. Granulation pattern, but not GSP or GHR mutation, is associated with clinical characteristics in somatostatin-naive patients with somatotroph adenomas. *Eur J Endocrinol.* 2013;168:491-9.
 28. Ibanez-Costa A, Korbonits M. AIP and the somatostatin system in pituitary tumours. *J Endocrinol.* 2017;235:R101-R116.
 29. Gadelha MR, Kasuki L, Korbonits M. The genetic background of acromegaly. *Pituitary.* 2017;20:10-21.
 30. Taboada GF, Neto LV, Luque RM. Impact of gsp oncogene on the mRNA content for somatostatin and dopamine receptors in human somatotropinomas. *Neuroendocrinology.* 2011;93:40-47.
 31. Luque RM, Ibáñez-Costa A, Neto LV. Truncated somatostatin receptor variant sst5TMD4 confers aggressive features (proliferation, invasion and reduced octreotide response) to somatotropinomas. *Cancer Lett.* 2015;359:299-306.
 32. Gatto F, Feelders RA, Franck SE. *In vitro* head-to-head comparison between octreotide and pasireotide in gh-secreting pituitary adenomas. *J Clin Endocrinol Metab.* 2017;102:2009-18.
 33. Chahal HS, Trivellin G, Leontiou CA. Somatostatin analogs modulate AIP in somatotroph adenomas: The role of the ZAC1 pathway. *J Clin Endocrinol Metab.* 2012;97:E1411-E1420.
 34. Theodoropoulou M, Stalla GK, Spengler D. ZAC1 target genes and pituitary tumorigenesis. *Mol Cell Endocrinol.* 2010;326:60-65.
 35. Garcia-Rendueles AR, Chenlo M, Oroz-Gonjar F. RET signaling provides tumorigenic mechanism and tissue specificity for AIP-related somatotrophinomas. *Oncogene.* 2021;40:6354-68.
 36. Tuominen I, Heliövaara E, Raitila A. AIP inactivation leads to pituitary tumorigenesis through defective Galphai-cAMP signaling. *Oncogene.* 2015;34:1174-84.
 37. Bogner EM, Daly AF, Gulde S. miR-34a is upregulated in AIP mutated somatotropinomas and promotes octreotide resistance. *Int J Cancer.* 2020;147:3523-38.
 38. Denes J, Kasuki L, Trivellin G. Regulation of aryl hydrocarbon receptor interacting protein (AIP) protein expression by MiR-34a in sporadic somatotropinomas. *PLoS One.* 2015;10:e0117107.
 39. Barry S, Carlsen E, Marques P. Tumor microenvironment defines the invasive phenotype of AIP-mutation-positive pituitary tumors. *Oncogene.* 2019;38:5381-95.
 40. Boguslowska A, Korbonits M. Genetics of acromegaly and gigantism. *J Clin Med.* 2021;10:1377.
 41. Leontiou CA, Gueorguiev M, van der Spuy J. The role of the aryl hydrocarbon receptor-interacting protein gene in familial and sporadic pituitary adenomas. *J Clin Endocrinol Metab.* 2008;93:2390-401.
 42. Trivellin G, Daly AF, Fauz FR. Gigantism and acromegaly due to Xq26 microduplications and GPR101 mutation. *N Engl J Med.* 2014;371:2363-74.
 43. Iacovazzo D, Caswell R, Bunce B. Germline or somatic GPR101 duplication leads to X-linked acrogigantism: A clinicopathological and genetic study. *Acta Neuropathol Commun.* 2016;4:56.
 44. Beckers A, Lodish MB, Trivellin G. X-linked acrogigantism syndrome: clinical profile and therapeutic responses. *Endocr Relat Cancer.* 2015;22:353-67.
 45. Vasilev V, Daly AF, Trivellin G, Stratakis CA, Zacharieva S, Beckers A. Hereditary endocrine tumors: Current state-of-the-art and research opportunities: The roles of AIP and GPR101 in Familial Isolated Pituitary Adenomas (FIPA). *Endocr Relat Cancer.* 2020;27:T77-T86.
 46. Daly AF, Yuan B, Fina F. Somatic mosaicism underlies X-linked acrogigantism syndrome in sporadic male subjects. *Endocr Relat Cancer.* 2016;23:221-33.
 47. Rodd C, Millette M, Iacovazzo D. Somatic GPR101 duplication causing X-Linked Acrogigantism (XLAG)-diagnosis and management. *J Clin Endocrinol Metab.* 2016;101:1927-30.
 48. Kamenicky P, Bouligand J, Chanson P. Gigantism, acromegaly, and GPR101 mutations. *N Engl J Med.* 2015;372:1264.
 49. Daly AF, Lysy PA, Desfilles C. GHRH excess and blockade in X-LAG syndrome. *Endocr Relat Cancer.* 2016;23:161-70.
 50. Iacovazzo D, Korbonits M. Gigantism: X-linked acrogigantism and GPR101 mutations. *Growth Horm IGF Res.* 2016;30:31-64-9.
 51. Hernández-Ramírez LC, Gam R, Valdés N. Loss-of-function mutations in the CABLES1 gene are a novel cause of Cushing's disease. *Endocr Relat Cancer.* 2017;24:379-92.
 52. Roussel-Gervais A, Couture C, Langlais D. The Cables1 gene in glucocorticoid regulation of pituitary corticotrope growth and Cushing disease. *J Clin Endocrinol Metab.* 2016;101:513-22.
 53. Zhang Q, Peng C, Song J. Germline mutations in CDH23, encoding cadherin-related 23, are associated with both familial and sporadic pituitary adenomas. *Am J Hum Genet.* 2017;100:817-23.
 54. Bolz H, von Brederlow B, Ramírez A. Mutation of CDH23, encoding a new member of the cadherin gene family, causes Usher syndrome type 1D. *Nat Genet.* 2001;27:108-12.
 55. Thakker RV. Multiple Endocrine Neoplasia type 1 (MEN1) and type 4 (MEN4). *Mol Cell Endocrinol.* 2014;386:2-15.
 56. Scherthaner-Reiter MH, Trivellin G, Stratakis CA. MEN1, MEN4, and carney complex: Pathology and molecular genetics. *Neuroendocrinology.* 2016;103:18-31.
 57. Chandrasekharappa SC, Guru SC, Manickam P. Positional cloning of the gene for multiple endocrine neoplasia-type 1. *Science.* 1997;276:404-7.
 58. Lemmens I. Identification of the Multiple Endocrine Neoplasia type 1 (MEN1) gene. The European Consortium on MEN1. *Hum Mol Genet.* 1997;6:1177-83.
 59. Machens A, Schaaf L, Karges W. Age-related penetrance of endocrine tumours in Multiple Endocrine Neoplasia type 1 (MEN1): A multicentre study of 258 gene carriers. *Clin Endocrinol (Oxf).* 2007;67:613-22.
 60. Agarwal SK. The future: Genetics advances in MEN1 therapeutic approaches and management strategies. *Endocr Relat Cancer.* 2017;24:T119-T134.

61. de Laat JM, Dekkers OM, Pieterman CR. Long-term natural course of pituitary tumors in patients with MEN1: Results from the Dutch MEN1 study group (DMSG). *J Clin Endocrinol Metab.* 2015;100:3288-96.
62. Vergès B, Boureille F, Goudet P. Pituitary disease in MEN type 1 (MEN1): Data from the France-Belgium MEN1 multicenter study. *J Clin Endocrinol Metab.* 2002;87:457-65.
63. Trouillas J, Labat-Moleur F, Sturm N. Pituitary tumors and hyperplasia in multiple endocrine neoplasia type 1 syndrome (MEN1): A case-control study in a series of 77 patients versus 2509 non-MEN1 patients. *Am J Surg Pathol.* 2008;32:534-43.
64. Borson-Chazot F, Garby L, Raverot G. Acromegaly induced by ectopic secretion of GHRH: A review 30 years after GHRH discovery. *Ann Endocrinol (Paris).* 2012;73:497-502.
65. Srirangam Nadhamuni V, Iacovazzo D, Evanson J. GHRH secretion from a pancreatic neuroendocrine tumor causing gigantism in a patient with MEN1. *Endocrinol Diabetes Metab Case Rep.* 2021;2021:1-8.
66. Kato M, Inoshita N, Sugiyama T. Differential expression of genes related to drug responsiveness between sparsely and densely granulated somatotroph adenomas. *Endocr J.* 2012;59:221-8.
67. Kiseljak-Vassiliades K, Xu M, Mills TS. Differential Somatostatin Receptor (SSTR) 1-5 expression and downstream effectors in histologic subtypes of growth hormone pituitary tumors. *Mol Cell Endocrinol.* 2015;417:73-83.
68. Alrezk R, Hannah-Shmouni F, Stratakis CA. MEN4 and CDKN1B mutations: The latest of the MEN syndromes. *Endocr Relat Cancer.* 2017;24:T195-T208.
69. Frederiksen A, Rossing M, Hermann P, Ejersted C, Thakker RV, Frost M. Clinical features of multiple endocrine neoplasia type 4: Novel pathogenic variant and review of published cases. *J Clin Endocrinol Metab.* 2019;104:3637-46.
70. Sambugaro S, Di Ruvo M, Ambrosio MR. Early onset acromegaly associated with a novel deletion in CDKN1B 5'UTR region. *Endocrine.* 2015;49:58-64.
71. Chevalier B, Odou MF, Demonchy J, Cardot-Bauters C, Vantyghem MC. Multiple endocrine neoplasia type 4: Novel CDKN1B variant and immune anomalies. *Ann Endocrinol (Paris).* 2020;81:124-5.
72. Fritz A, Walch A, Piotrowska K. Recessive transmission of a multiple endocrine neoplasia syndrome in the rat. *Cancer Res.* 2002;62:3048-51.
73. Pellegata NS, Quintanilla-Martinez L, Siggelkow H. Germ-line mutations in p27Kip1 cause a multiple endocrine neoplasia syndrome in rats and humans. *Proc Natl Acad Sci USA.* 2006;103:15558-63.
74. Agarwal SK, Mateo CM, Marx SJ. Rare germline mutations in cyclin-dependent kinase inhibitor genes in multiple endocrine neoplasia type 1 and related states. *J Clin Endocrinol Metab.* 2009;94:1826-34.
75. Chasseloup F, Pankratz N, Lane J. Germline CDKN1B loss-of function variants cause pediatric Cushing's disease with or without an MEN4 phenotype. *J Clin Endocrinol Metab.* 2020;105:1983-2005.
76. Stratakis CA, Kirschner LS, Carney JA. Clinical and molecular features of the Carney complex: Diagnostic criteria and recommendations for patient evaluation. *J Clin Endocrinol Metab.* 2001;86:4041-6.
77. Carney JA, Gordon H, Carpenter PC, Shenoy BV, Go VL. The complex of myxomas, spotty pigmentation, and endocrine overactivity. *Medicine (Baltimore).* 1985;64:270-83.
78. Bertherat J, Horvath A, Groussin L. Mutations in regulatory subunit type 1A of cyclic adenosine 5'-monophosphate-dependent protein kinase (PRKAR1A): Phenotype analysis in 353 patients and 80 different genotypes. *J Clin Endocrinol Metab.* 2009;94:2085-91.
79. Correa R, Salpea P, Stratakis CA. Carney complex: An update. *Eur J Endocrinol.* 2015;173:M85-M97.
80. Salpea P, Stratakis CA. Carney complex and McCune Albright syndrome: An overview of clinical manifestations and human molecular genetics. *Mol Cell Endocrinol.* 2014;386:85-91.
81. Stelmachowska-Banas M, Zgliczynski W, Tutka P, Carney JA, Korbonits M. Fatal Carney complex in siblings due to *de novo* large gene deletion. *J Clin Endocrinol Metab.* 2017;102:3924-7.
82. Kirschner LS, Carney JA, Pack SD. Mutations of the gene encoding the protein kinase A type I-alpha regulatory subunit in patients with the Carney complex. *Nat Genet.* 2000;26:89-92.
83. Caimari F, Korbonits M. Novel genetic causes of pituitary adenomas. *Clin Cancer Res.* 2016;22:5030-42.
84. Kiefer FW, Winhofer Y, Iacovazzo D. PRKAR1A mutation causing pituitary-dependent Cushing disease in a patient with Carney complex. *Eur J Endocrinol.* 2017;177:K7-K12.
85. Hernández-Ramírez LC, Tatsi C, Lodish MB. Corticotropinoma as a component of Carney complex. *J Endocr Soc.* 2017;1:918-25.
86. Dénes J, Swords F, Rattenberry E. Heterogeneous genetic background of the association of pheochromocytoma/paraganglioma and pituitary adenoma: Results from a large patient cohort. *J Clin Endocrinol Metab.* 2015;100:E531-E541.
87. Gill AJ, Toon CW, Clarkson A. Succinate dehydrogenase deficiency is rare in pituitary adenomas. *Am J Surg Pathol.* 2014;38:560-6.
88. Pozza C, Sesti F, Di Dato C. A novel MAX gene mutation variant in a patient with multiple and "composite" neuroendocrine-neuroblastic tumors. *Front Endocrinol (Lausanne).* 2020;11:234.
89. Schultz KAP, Williams GM, Kamihara J. DICER1 and associated conditions: Identification of at-risk individuals and recommended surveillance strategies. *Clin Cancer Res.* 2018;24:2251-61.
90. Doros LA, Rossi CT, Yang J. DICER1 mutations in childhood cystic nephroma and its relationship to DICER1-renal sarcoma. *Mod Pathol.* 2014;27:1267-80.
91. de Kock L, Priest JR, Foulkes WD, Alexandrescu S. An update on the central nervous system manifestations of DICER1 syndrome. *Acta Neuropathol.* 2020;139:689-701.
92. Scheithauer BW, Horvath E, Abel TW. Pituitary blastoma: A unique embryonal tumor. *Pituitary blastoma: A unique embryonal tumor. Pituitary.* 2012;15:365-73.
93. Cotton E, Ray D. DICER1 mutation and pituitary prolactinoma. *Endocrinol Diabetes Metab Case Rep.* 2018;2018:1-4.
94. Choong CS, Priest JR, Foulkes WD. Exploring the endocrine manifestations of DICER1 mutations. *Trends Mol Med.* 2012;18:503-5.
95. Krol J, Loedige I, Filipowicz W. The widespread regulation of microRNA biogenesis, function and decay. *Nat Rev Genet.* 2010;11:597-610.
96. Uraki S, Ariyasu H, Doi A. Atypical pituitary adenoma with MEN1 somatic mutation associated with abnormalities of DNA mismatch repair genes; MLH1 germline mutation and MSH6 somatic mutation. *Endocr J.* 2017;64:895-906.
97. Teuber J, Reinhardt A, Reuss D, Hahnel S, Unterberg A, Beynon C. Aggressive pituitary adenoma in the context of Lynch syndrome: A case report and literature review on this rare coincidence. *Br J Neurosurg.* 2021;35:1-6.
98. Tadini G, Milani D, Menni F, Pezzani L, Sabatini C, Esposito S. Is it time to change the neurofibromatosis 1 diagnostic criteria? *Eur J Intern Med.* 2014;25:506-10.
99. Milani D, Pezzani L, Tadini G, Menni F, Esposito S. A multidisciplinary approach in neurofibromatosis 1. *Lancet Neurol.* 2015;14:29-30.
100. Rosner M, Hanneder M, Siegel N, Valli A, Fuchs C, Hengstschlager M. The mTOR pathway and its role in human genetic diseases. *Mutat Res.* 2008;659:284-92.